

# Optimizing Tissue Culture Methods in Diverse Nightshade Species

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## Introduction

- In vitro* propagation is essential for the generation of stable transgenic plant lines in *Solanaceae* and for propagation of sterile lines.
- We are interested studying fruit evolution by creating transgenic knockout lines in nightshade species with diverse fruit types.
- We have adapted a tomato (*Solanum lycopersicum*) *in vitro* tissue culture and transformation protocol from the Boyce Thompson institute for use in a diverse group of *Solanaceae* species with useful phylogenetic placement:
  - Fleshy Fruited Species:
    - Wild tomato (*Solanum pimpinellifolium*)
    - Cestrum spp.* & *Brunfelsia*, fleshy-fruited species in the dry-fruited grade
  - Dry-fruited Species:
    - Synthetic tobacco (*N. tabacum*) hybrids
    - Desert tobacco (*Nicotiana obtusifolia*), a small diploid congener of *N. tabacum* more amenable to greenhouse growth conditions
    - Jimsonweed (*Datura stramonium*), a dry-fruited species in the fleshy *Solanoideae* clade
    - Schizanthus grahamii*, a very early diverging species within *Solanaceae*

## Results

### Desert Tobacco (*Nicotiana obtusifolia*)

| Callus Induction            | Shoot Induction  | Root Induction                  | Agro Transformation           | Transfer to Soil |
|-----------------------------|--|---------------------------------|-------------------------------|------------------|
|                             |  |                                 |                               |                  |
| Cotyledons<br>0.2mg/L 2,4-D | 2 <sup>mg</sup> /L zeatin then 1 <sup>mg</sup> /L zeatin | 0.5-1.0 <sup>mg</sup> /L zeatin | Success with kan <sup>R</sup> | Success          |

### Jimsonweed (*D. stramonium*)

| Callus Induction            | Shoot Induction  | Root Induction         | Agro Transformation | Transfer to Soil |
|-----------------------------|--|------------------------|---------------------|------------------|
|                             |  |                        |                     |                  |
| Cotyledons<br>0.2mg/L 2,4-D | 2 <sup>mg</sup> /L zeatin then 1 <sup>mg</sup> /L zeatin | 1 <sup>mg</sup> /L IAA | Success with erGFP  | Success          |

### Schizanthus

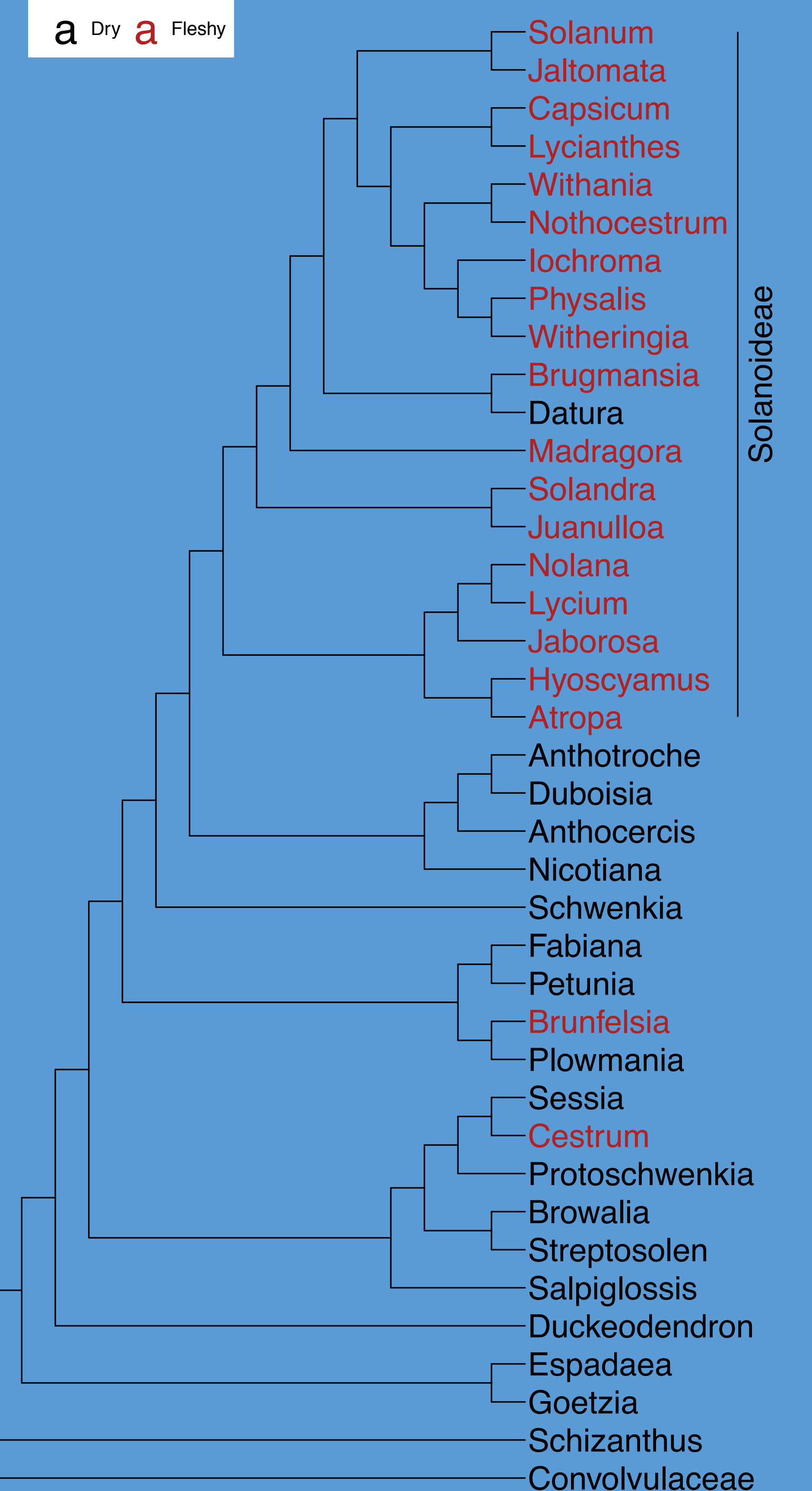
| Callus Induction            | Shoot Induction                  |
|-----------------------------|----------------------------------|
|                             |                                  |
| Cotyledons<br>0.2mg/L 2,4-D | 2mg/L zeatin then 0.5mg/L zeatin |

### Brunfelsia

| Callus Induction           |
|----------------------------|
|                            |
| Leaf Disc<br>0.2mg/L 2,4-D |

### Cestrum

| Callus Induction           |
|----------------------------|
|                            |
| Leaf Disc<br>0.2mg/L 2,4-D |



## Methods

All explants were grown on MS media supplemented with Nitsch vitamins in 16h light at 22°C. Filter-sterilized plant growth regulators including zeatin and indole-3-acetic acid were added at the concentrations indicated after autoclaving. Initial explants were grown on 15mm-deep petri dishes; after the establishment of callus, they were transferred to 16oz plastic containers.

## Conclusions and Future Work

- Viable *N. obtusifolia*, *Nicotiana* hybrids, and *D. stramonium* have been regenerated from callus.
- Schizanthus*, *Cestrum*, and *Brunfelsia* are still in progress, but show promise.

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